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LETTERS

# New stereocontrolled transformations in the imidazolosugar series 

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#### Abstract

The isomeric imidazolopyrrolidinose 1, imidazolopiperidinose 2 and imidazoloazepanose 3, potential glycosidase inhibitors, were obtained in several steps from D-glucose. © 2002 Elsevier Science Ltd. All rights reserved.


Imidazolosugars as potential glycosidase inhibitors have been a subject of interest for several research groups. ${ }^{1-6}$ The only known natural imidazolosugarnagstatine is an effective inhibitor of $N$-acetylamino- $\beta$ -D-glucosaminidase of bovine kidney. ${ }^{7,8}$ We present herein the synthesis of three new isomeric imidazolosugars of the structures 1, 2 and 3 (Fig. 1), potential glycosidase inhibitors.

This synthesis is based on stereocontrolled transformations of two epimeric dibenzylditritylimidazolyl-pentitols 10a and 10b which were obtained in several steps from dialdofuranose 4, a compound readily available from D-glucose. ${ }^{9}$

The two epimeric imidazolosugars $\mathbf{6 a}$ and $\mathbf{6 b}$ were prepared in a $3: 5$ ratio by nucleophilic addition of the 5-lithiated derivative of $2-t$-butyldimethylsilyl-1dimethylsulphamoylimidazole (5) ${ }^{10}$ to dialdofuranose $4^{9}$ according to the Kurihara procedure. ${ }^{11}$ These two diastereomers were separated by flash chromatography and hence subjected to analogous reaction sequences.


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Thus, benzylation of $\mathbf{6 a}$ and $\mathbf{6 b}$ resulted in the formation of the compounds 7a and 7b which, after removing acid-labile protecting groups, gave imidazolyl-pentoses $\mathbf{8 a}$ and $\mathbf{8 b}$, each as a mixture of anomers. Subsequent reduction of $\mathbf{8 a}$ and $\mathbf{8 b}$ yielded imidazolyl-pentitols $\mathbf{9 a}$ and 9b, respectively, which were then tritylated to form the ditrityl derivatives 10a and 10b (Scheme 1).

Phenylmethanesulphonylation of the OH groups in 10a and successive removal of the trityl groups by acid hydrolysis afforded the imidazolyl-pentitol 11a. The same reaction sequence starting from 10b gave the imidazolyl-pentitol 11b and the imidazolo-manno-piperidinose $\mathbf{1 2}$ in a $2: 1$ ratio (Scheme 2).

Under treatment with $\mathrm{NaNH}_{2}$ the compound 11a rearranged to the tricyclic structure 13, ${ }^{14}$ which was consecutively catalytically debenzylated and the resulting aldehyde was immediately reduced to produce the imidazolopyrrolidinose 1. ${ }^{14}$ Imidazolopiperidinose 12, treated with $\mathrm{NaNH}_{2}$, underwent a trans-elimination reaction to provide its unsaturated derivative 14, ${ }^{14}$ which was catalytically debenzylated and reduced to give the imidazolopiperidinose $2 .{ }^{14}$ When the imida-zolyl-pentitol 11b was treated with $\mathrm{NaNH}_{2}$, the cyclisation and elimination reaction sequence led to the dibenzyl unsaturated imidazoloazepanose 15. ${ }^{14}$ Catalytic debenzylation and reduction of $\mathbf{1 5}$ resulted in the formation of the imidazoloazepanose $3^{14}$ (Scheme 3).

The stereochemical outcome of the above reactions proves the configurations at the newly occurring chiral centre of the two epimeric adducts $\mathbf{6 a}$ and $\mathbf{6 b}$. Thus, we believe the strong base promotes the $\mathrm{S}_{\mathrm{N}} 2$ type cyclisation of 11a to afford the imidazolopiperidinose 16

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Scheme 1. Reagents and conditions: (a) 2-t-butyldimethylsilyl-1-dimethylsulphamoylimidazole (5), BuLi, THF, $-70^{\circ} \mathrm{C}, 78 \%$; (b) $\mathrm{BnBr}, \mathrm{NaH}, \mathrm{DMF}$, rt; (c) $1.5 \mathrm{~N} \mathrm{HCl}, \mathrm{THF}$, reflux, 8a: $65 \%$ (two steps), $\mathbf{8 b}: 60 \%$ (two steps); (d) $\mathrm{NaBH}_{4}, \mathrm{MeOH}, 0^{\circ} \mathrm{C} \rightarrow \mathrm{rt}, \mathbf{9 a}: 88 \%$, 9b: $87 \%$; (e) $\mathrm{TrCl}, \mathrm{Py}, \mathrm{DMAP}, 80^{\circ} \mathrm{C}, 10 \mathrm{a}: 56 \%$, 10b: $48 \%$.


Scheme 2. Reagents and conditions: (a) $\mathrm{BnSO}_{2} \mathrm{Cl}, \mathrm{Py}$, $-30^{\circ} \mathrm{C} \rightarrow \mathrm{rt}$; (b) $6 \mathrm{~N} \mathrm{HCl}, \mathrm{THF}$, reflux, 11a: $70 \%$ (two steps); 11b: $34 \%$, 12: $18 \%$ (two steps).
(Scheme 4). The rearrangement of $\mathbf{1 6}$ into $\mathbf{1 3}$ involves piperidine ring contraction, relying on the departure of the equatorial phenylmethanesulphonate group from C-7 and the 1,2-migration of the antiperiplanar [C-8/C8a] bond, assisted by an attack of the alcoholate ion at C-9 on C-8, which is possible only in the boat conformation of the piperidine ring. The formation of $\mathbf{1 3}$ as only one isomer with the $S$-configuration at C-8 suggests both the above processes are simultaneous (Scheme 4). The epimeric imidazolopiperidinose 12, however, in the same reaction conditions yields the trans-elimination product 14 , thanks to the hydrogen atom at C-8 in $\mathbf{1 2}$ which is situated anti towards the leaving group.

The conversion of the imidazolyl-pentitol 11b into the imidazoloazepanose 15 in basic conditions proceeds probably via the epoxide 17 (compare Lohray et al. ${ }^{13}$ ), with seven-membered ring closing and trans-elimina-





Scheme 3. Reagents and conditions: (a) $\mathrm{NaNH}_{2}, \mathrm{DMF}, \mathrm{rt}, 13$ : $56 \%$, 14: $84 \%, 15: 85 \%$; (b) $\mathrm{H}_{2} / \mathrm{Pd} / \mathrm{C}, \mathrm{EtOH}$, rt, 1: $81 \%, 2$ : $35 \%$, 3: $24 \%$.
tion of phenylmethanesulphonic acid residue (Scheme 5).

The trans stereochemistry of the elimination reactions converting $\mathbf{1 2}$ into $\mathbf{1 4}$ and $\mathbf{1 1 b}$ into $\mathbf{1 5}$ proves the $R$-configuration on the carbon atom adjacent to the imidazole ring in both $\mathbf{1 2}$ and 11b.

The configuration of $\mathbf{1 3}$ and the boat conformation of its pyranose ring (Scheme 3) were deduced by nuclear Overhauser effect (NOE) measurements. Thus, irradiation of H-9 generated nuclear Overhauser enhancement at H-9' $(26 \%)$ and $\mathrm{H}-5(6 \%)$. When H-9' was irradiated, a $22 \%$ NOE at $\mathrm{H}-9,8 \%$ NOE at $\mathrm{H}-8$ and $4 \%$ at $\mathrm{H}-5$ were observed. Irradiation of H-7 gave enhancement of H-8 ( $3 \%$ ) and H-6 (8\%). The irradiation of H-6 led only to NOE enhancement of the proton H-7 (10\%). The experimental results were confirmed by molecular modelling with the SYBYL 6.5 software from TRIPOS Inc.


16
Scheme 4. Proposed course of rearrangement of $\mathbf{1 6}$ into 13.


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Scheme 5. Proposed course of the seven-membered ring closing in 17.

The configuration at $\mathrm{C}-8$ in compound 2 was assigned on the basis of the ROE signal between H-6 and H-8 in the ROESY spectrum. The ROE signal between $\mathrm{H}-7$ and $\mathrm{H}-9$ was helpful in the configuration assignment at C-9 for compound 3.

The configuration at C-6 in 3 can be deduced from the analysis of vicinal coupling constants. The proton H-6 occupies the pseudo-equatorial position, corresponding to the $R$-configuration, because its vicinal coupling constants with the pseudo-axial H-5 and H-7 (0 and 1.75 Hz , respectively) correspond to $\mathrm{H}-\mathrm{C}-\mathrm{C}-\mathrm{H}$ torsion angles close to $90^{\circ} .{ }^{12}$

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14. All new compounds gave satisfactory NMR and MS spectroscopy data. The most important data are listed below.
1: $[\alpha]_{\mathrm{D}}-16(c \quad 0.3, \mathrm{MeOH}) ;{ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{D}_{2} \mathrm{O}\right) \delta: 3.43$ (ddd, $1 \mathrm{H}, J 5.1,6.2,6.6 \mathrm{~Hz}, \mathrm{H}-5), 3.85(\mathrm{dd}, 1 \mathrm{H}, J 6.2,11.3 \mathrm{~Hz}$, H-9), 3.90 (dd, $1 \mathrm{H}, J 6.6,12.4 \mathrm{~Hz}, \mathrm{H}-8$ ), 4.00 (dd, $1 \mathrm{H}, J$ $\left.5.1,11.3 \mathrm{~Hz}, \mathrm{H}-9^{\prime}\right), 4.22$ (dd, 1H, J 3.7, $12.4 \mathrm{~Hz}, \mathrm{H}^{\prime} 8^{\prime}$ ), $4.50(\mathrm{dt}, 1 \mathrm{H}, J 3.7,6.6 \mathrm{~Hz}, \mathrm{H}-7), 4.61(1 \mathrm{H}, \mathrm{t}, J 6.6 \mathrm{~Hz}$, $\mathrm{H}-6), 7.24$ ( $\mathrm{s}, 1 \mathrm{H}, \mathrm{H}-1$ ), 8.61 (s, $1 \mathrm{H}, \mathrm{H}-3$ ); ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{D}_{2} \mathrm{O}\right) \delta: 45.0(\mathrm{C}-5), 58.2(\mathrm{C}-8), 58.3(\mathrm{C}-9), 65.5(\mathrm{C}-7)$, 75.0 (C-6), 111.6 (C-1), 127.0 (C-3), 134.6 (C-7a); FAB MS: $185\left([\mathrm{M}+\mathrm{H}]^{+}\right)$; HRMS: calcd for $[\mathrm{M}+\mathrm{H}]^{+}$ $\left(\mathrm{C}_{8} \mathrm{H}_{13} \mathrm{~N}_{2} \mathrm{O}_{3}\right) 185.0926$, found 185.0926
2: $[\alpha]_{\mathrm{D}}-32(c 0.35, \mathrm{MeOH}) ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{D}_{2} \mathrm{O}\right) \delta: 1.86$ (ddd, 1H, J 7.5, 8.75, $13.0 \mathrm{~Hz}, \mathrm{H}-7$ ), 2.32 (ddd, 1H, J 3.0, $5.75,13.0 \mathrm{~Hz}, \mathrm{H}-7$ '), 3.78 (dd, 1H, J $4.25,12.25 \mathrm{~Hz}, \mathrm{H}-9$ ), 3.97 (dd, 1H, J 3.5, $12.25 \mathrm{~Hz}, \mathrm{H}^{\prime} 9^{\prime}$ ), 4.03 (ddd, 1H, J 3.5, $4.25,6.5 \mathrm{~Hz}, \mathrm{H}-5), 4.12$ (ddd, $1 \mathrm{H}, J 3.0,6.5,8.75 \mathrm{~Hz}$, $\mathrm{H}-6), 4.89(1 \mathrm{H}, \mathrm{dd}, J 5.75,7.5 \mathrm{~Hz}, \mathrm{H}-8), 6.95(\mathrm{~s}, 1 \mathrm{H}$, $\mathrm{H}-1), 7.73$ (s, $1 \mathrm{H}, \mathrm{H}-3$ ); ${ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{D}_{2} \mathrm{O}\right) \delta: 35.9(\mathrm{C}-7)$, 60.2 (C-9), 60.3 (C-8), 61.0 (C-5), 64.0 (C-6), 124.0 (C-1), 131.2 (C-3), 143.0 (C-8a); FAB MS: $185\left([\mathrm{M}+\mathrm{H}]^{+}\right)$; HRMS: calcd for $[\mathrm{M}+\mathrm{H}]^{+}\left(\mathrm{C}_{8} \mathrm{H}_{13} \mathrm{~N}_{2} \mathrm{O}_{3}\right)$ 185.0926, found 185.0923

3: $[\alpha]_{\mathrm{D}}-17(c 0.4, \mathrm{MeOH}) ;{ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{D}_{2} \mathrm{O}\right) \delta: 2.03$ (ddd, $1 \mathrm{H}, J 10.5,10.75,12.5 \mathrm{~Hz}, \mathrm{H}-8$ ), 2.14 (ddd, $1 \mathrm{H}, J 3.0$, $\left.4.0,12.5 \mathrm{~Hz}, \mathrm{H}-8^{\prime}\right), 3.99(\mathrm{~d}, 1 \mathrm{H}, J 15.0 \mathrm{~Hz}, \mathrm{H}-5), 4.06$ (ddd, $1 \mathrm{H}, J 1.75,4.0,10.75 \mathrm{~Hz}, \mathrm{H}-7), 4.15$ (dd, $1 \mathrm{H}, J$ $1.75,6.5 \mathrm{~Hz}, \mathrm{H}-6), 4.37$ (dd, $\left.1 \mathrm{H}, J 6.5,15.0 \mathrm{~Hz}, \mathrm{H}^{\prime} 5^{\prime}\right)$, $4.89(\mathrm{dd}, 1 \mathrm{H}, J 3.0,10.5 \mathrm{~Hz}, \mathrm{H}-9), 6.95(\mathrm{~s}, 1 \mathrm{H}, \mathrm{H}-1), 7.73$ (s, $1 \mathrm{H}, \mathrm{H}-3) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{D}_{2} \mathrm{O}\right) \delta: 35.8(\mathrm{C}-8), 45.3(\mathrm{C}-5)$, 61.6 (C-9), 66.9 (C-7), 69.4 (C-6), 120.6 (C-1), 126.8 (C-3), 134.1 (C-9a); FAB MS: 185 ( $\left.[\mathrm{M}+\mathrm{H}]^{+}\right)$; HRMS: calcd for $[\mathrm{M}+\mathrm{H}]^{+}\left(\mathrm{C}_{8} \mathrm{H}_{13} \mathrm{~N}_{2} \mathrm{O}_{3}\right)$ 185.0926, found 185.0929 13: $[\alpha]_{\mathrm{D}}-82\left(c 1.75, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta: 3.48$ (bd, 1H, J $2.5 \mathrm{~Hz}, \mathrm{H}-7$ ), 3.60 (dd, 1H, J $3.25,11.25 \mathrm{~Hz}$, H-9), 3.85 (d, 1H, J $\left.11.25 \mathrm{~Hz}, \mathrm{H}^{\prime} 9^{\prime}\right), 4.40(\mathrm{~d}, 1 \mathrm{H}, J 3.25$ $\mathrm{Hz}, \mathrm{H}-5), 4.51,4.73$ (2d, 2H, J $12 \mathrm{~Hz}, \mathrm{OBn}$ ), 4.54, 4.62 (2d, 2H, J $11.5 \mathrm{~Hz}, \mathrm{OBn}), 4.88(\mathrm{~d}, 1 \mathrm{H}, J 2.5 \mathrm{~Hz}, \mathrm{H}-8)$, $4.89(\mathrm{~s}, 1 \mathrm{H}, \mathrm{H}-6), 6.86(\mathrm{~s}, 1 \mathrm{H}, \mathrm{H}-1), 7.19-7.42(\mathrm{~m}, 10 \mathrm{H}$, arom.), 7.51 ( $\mathrm{s}, 1 \mathrm{H}, \mathrm{H}-3$ ); ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta: 43.5$ (C-7), $59.2(\mathrm{C}-5), 63.5(\mathrm{C}-9), 69.5\left(\mathrm{OCH}_{2} \mathrm{Ph}\right), 70.7$ $\left(\mathrm{OCH}_{2} \mathrm{Ph}\right), 85.2(\mathrm{C}-6), 98.6$ (C-8), 121.2 (C-1), $128.0-$ 129.0 (m, arom.), 131.1 (C-3), 135.2 (C-7a), 138.8, 138.9 (s-arom.); FAB MS: 363 ([M+H] ${ }^{+}$)

14: $[\alpha]_{\mathrm{D}}+172\left(c \quad 0.9, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta$ : 3.47 (dd, 1H, J 8.75, $11.25 \mathrm{~Hz}, \mathrm{H}-9$ ), 3.60 (dd, $1 \mathrm{H}, J$ $\left.5.0,11.25 \mathrm{~Hz}, \mathrm{H}-9^{\prime}\right), 4.24$ (dd, $1 \mathrm{H}, J 1.25,6.25 \mathrm{~Hz}$, H-6), 4.41 (ddd, 1H, J $1.25,5.0,8.75 \mathrm{~Hz}, \mathrm{H}-5$ ), 4.42, $4.47(2 \mathrm{~d}, 2 \mathrm{H}, J 12.25 \mathrm{~Hz}, \mathrm{OBn}), 4.88(\mathrm{~d}, 1 \mathrm{H}, J 6.25$, $\mathrm{H}-7), 4.95(\mathrm{~s}, 2 \mathrm{H}, \mathrm{OBn}), 6.94(\mathrm{~s}, 1 \mathrm{H}, \mathrm{H}-1), 7.20-7.40$ (m, 10 H , arom.), $7.50(\mathrm{~s}, 1 \mathrm{H}, \mathrm{H}-3) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right)$ $\delta: 60.5 \quad(\mathrm{C}-5), \quad 62.8 \quad(\mathrm{C}-9) \quad 69.2 \quad\left(\mathrm{OCH}_{2} \mathrm{Ph}\right), \quad 69.3$ $\left(\mathrm{OCH}_{2} \mathrm{Ph}\right), 71.2$ (C-6), 89.8 (C-7), 124.8 (C-1), 127.5129.0 (m, arom.), 138.3 (C-3), 139.2 (C-8a), 140.0 (C-8); FAB MS: $363\left([\mathrm{M}+\mathrm{H}]^{+}\right)$

15: $[\alpha]_{\mathrm{D}}+57\left(c\right.$ 1.5, $\left.\mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}+\mathrm{C}_{6} \mathrm{D}_{6}\right) \delta:$ 3.51 (dd, 1H, J 2.6, 13.2 Hz, H-5), 3.74 (ddd, 1H, J 2.6, $3.1,9.0 \mathrm{~Hz}, \mathrm{H}-6), 3.83$ (dd, 1H, J 9.0, $13.2 \mathrm{~Hz}, \mathrm{H}-5^{\prime}$ ), 3.88 (dd, 1H, J 3.1, $6.0 \mathrm{~Hz}, \mathrm{H}-7), 4.21,4.36$ (2d, 2H, J 11.5 Hz , OBn), 4.58, 4.61 (2d, 2H, $J 12 \mathrm{~Hz}, \mathrm{OBn}$ ), $4.70(\mathrm{~d}, 1 \mathrm{H}, J$ $6.0, \mathrm{H}-8), 7.00(\mathrm{~s}, 1 \mathrm{H}, \mathrm{H}-1), 7.00-7.30(\mathrm{~m}, 10 \mathrm{H}$, arom.), 7.46 $(\mathrm{s}, 1 \mathrm{H}, \mathrm{H}-3) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}+\mathrm{C}_{6} \mathrm{D}_{6}\right) \delta: 47.9(\mathrm{C}-5) 69.9$ $(\mathrm{C}-6), 70.3\left(\mathrm{OCH}_{2} \mathrm{Ph}\right), 71.2\left(\mathrm{OCH}_{2} \mathrm{Ph}\right), 76.4(\mathrm{C}-7), 95.8$ (C-8), 127.5-129.0 (m, arom.), 131.0 (C-3), 137.7, 139.4 (s-arom.), 138.0 (C-9a), 140.0 (C-1), 147.8 (C-9); FAB MS: 363 ( $\left.\mathrm{M}+\mathrm{H}]^{+}\right)$.


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