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New stereocontrolled transformations in the imidazolosugar series

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Abstract—The isomeric imidazolopyrrolidinose 1, imidazolopiperidinose 2 and imidazoloazepanose 3, potential glycosidase inhibitors, were obtained in several steps from D-glucose. © 2002 Elsevier Science Ltd. All rights reserved.

Imidazolosugars as potential glycosidase inhibitors have been a subject of interest for several research groups.¹⁻⁶ The only known natural imidazolosugar– nagstatine is an effective inhibitor of *N*-acetylamino- β -D-glucosaminidase of bovine kidney.^{7,8} We present herein the synthesis of three new isomeric imidazolosugars of the structures **1**, **2** and **3** (Fig. 1), potential glycosidase inhibitors.

This synthesis is based on stereocontrolled transformations of two epimeric dibenzylditritylimidazolyl-pentitols **10a** and **10b** which were obtained in several steps from dialdofuranose **4**, a compound readily available from D-glucose.⁹

The two epimeric imidazolosugars **6a** and **6b** were prepared in a 3:5 ratio by nucleophilic addition of the 5-lithiated derivative of 2-*t*-butyldimethylsilyl-1dimethylsulphamoylimidazole (**5**)¹⁰ to dialdofuranose **4**⁹ according to the Kurihara procedure.¹¹ These two diastereomers were separated by flash chromatography and hence subjected to analogous reaction sequences.



Figure 1.

Thus, benzylation of **6a** and **6b** resulted in the formation of the compounds **7a** and **7b** which, after removing acid-labile protecting groups, gave imidazolyl-pentoses **8a** and **8b**, each as a mixture of anomers. Subsequent reduction of **8a** and **8b** yielded imidazolyl-pentitols **9a** and **9b**, respectively, which were then tritylated to form the ditrityl derivatives **10a** and **10b** (Scheme 1).

Phenylmethanesulphonylation of the OH groups in 10a and successive removal of the trityl groups by acid hydrolysis afforded the imidazolyl-pentitol 11a. The same reaction sequence starting from 10b gave the imidazolyl-pentitol 11b and the imidazolo-*manno*-pipe-ridinose 12 in a 2:1 ratio (Scheme 2).

Under treatment with NaNH₂ the compound **11a** rearranged to the tricyclic structure **13**,¹⁴ which was consecutively catalytically debenzylated and the resulting aldehyde was immediately reduced to produce the imidazolopyrrolidinose **1**.¹⁴ Imidazolopiperidinose **12**, treated with NaNH₂, underwent a *trans*-elimination reaction to provide its unsaturated derivative **14**,¹⁴ which was catalytically debenzylated and reduced to give the imidazolopiperidinose **2**.¹⁴ When the imidazolyl-pentitol **11b** was treated with NaNH₂, the cyclisation and elimination reaction sequence led to the dibenzyl unsaturated imidazoloazepanose **15**.¹⁴ Catalytic debenzylation and reduction of **15** resulted in the formation of the imidazoloazepanose **3**¹⁴ (Scheme 3).

The stereochemical outcome of the above reactions proves the configurations at the newly occurring chiral centre of the two epimeric adducts **6a** and **6b**. Thus, we believe the strong base promotes the S_N^2 type cyclisation of **11a** to afford the imidazolopiperidinose **16**

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Scheme 1. Reagents and conditions: (a) 2-t-butyldimethylsilyl-1-dimethylsulphamoylimidazole (5), BuLi, THF, -70° C, 78%; (b) BnBr, NaH, DMF, rt; (c) 1.5N HCl, THF, reflux, 8a: 65% (two steps), 8b: 60% (two steps); (d) NaBH₄, MeOH, 0°C \rightarrow rt, 9a: 88%, 9b: 87%; (e) TrCl, Py, DMAP, 80°C, 10a: 56%, 10b: 48%.



Scheme 2. Reagents and conditions: (a) $BnSO_2Cl$, Py, $-30^{\circ}C \rightarrow rt$; (b) 6N HCl, THF, reflux, 11a: 70% (two steps); 11b: 34%, 12: 18% (two steps).

(Scheme 4). The rearrangement of 16 into 13 involves piperidine ring contraction, relying on the departure of the equatorial phenylmethanesulphonate group from C-7 and the 1,2-migration of the antiperiplanar [C-8/C-8a] bond, assisted by an attack of the alcoholate ion at C-9 on C-8, which is possible only in the boat conformation of the piperidine ring. The formation of 13 as only one isomer with the S-configuration at C-8 suggests both the above processes are simultaneous (Scheme 4). The epimeric imidazolopiperidinose 12, however, in the same reaction conditions yields the *trans*-elimination product 14, thanks to the hydrogen atom at C-8 in 12 which is situated *anti* towards the leaving group.

The conversion of the imidazolyl-pentitol **11b** into the imidazoloazepanose **15** in basic conditions proceeds probably via the epoxide **17** (compare Lohray et al.¹³), with seven-membered ring closing and *trans*-elimina-



Scheme 3. *Reagents and conditions*: (a) NaNH₂, DMF, rt, 13: 56%, 14: 84%, 15: 85%; (b) H₂/Pd/C, EtOH, rt, 1: 81%, 2: 35%, 3: 24%.

tion of phenylmethanesulphonic acid residue (Scheme 5).

The *trans* stereochemistry of the elimination reactions converting 12 into 14 and 11b into 15 proves the R-configuration on the carbon atom adjacent to the imidazole ring in both 12 and 11b.

The configuration of 13 and the boat conformation of its pyranose ring (Scheme 3) were deduced by nuclear Overhauser effect (NOE) measurements. Thus, irradiation of H-9 generated nuclear Overhauser enhancement at H-9' (26%) and H-5 (6%). When H-9' was irradiated, a 22% NOE at H-9, 8% NOE at H-8 and 4% at H-5 were observed. Irradiation of H-7 gave enhancement of H-8 (3%) and H-6 (8%). The irradiation of H-6 led only to NOE enhancement of the proton H-7 (10%). The experimental results were confirmed by molecular modelling with the SYBYL 6.5 software from TRIPOS Inc.



Scheme 4. Proposed course of rearrangement of 16 into 13.



Scheme 5. Proposed course of the seven-membered ring closing in 17.

The configuration at C-8 in compound 2 was assigned on the basis of the ROE signal between H-6 and H-8 in the ROESY spectrum. The ROE signal between H-7 and H-9 was helpful in the configuration assignment at C-9 for compound 3.

The configuration at C-6 in **3** can be deduced from the analysis of vicinal coupling constants. The proton H-6 occupies the *pseudo*-equatorial position, corresponding to the *R*-configuration, because its vicinal coupling constants with the *pseudo*-axial H-5 and H-7 (0 and 1.75 Hz, respectively) correspond to H–C–C–H torsion angles close to 90° .¹²

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- All new compounds gave satisfactory NMR and MS spectroscopy data. The most important data are listed below.

1: $[\alpha]_{D}$ –16 (*c* 0.3, MeOH); ¹H NMR (D₂O) δ : 3.43 (ddd, 1H, *J* 5.1, 6.2, 6.6 Hz, H-5), 3.85 (dd, 1H, *J* 6.2, 11.3 Hz, H-9), 3.90 (dd, 1H, *J* 6.6, 12.4 Hz, H-8), 4.00 (dd, 1H, *J* 5.1, 11.3 Hz, H-9'), 4.22 (dd, 1H, *J* 3.7, 12.4 Hz, H-8'), 4.50 (dt, 1H, *J* 3.7, 6.6 Hz, H-7), 4.61 (1H, t, *J* 6.6 Hz, H-6), 7.24 (s, 1H, H-1), 8.61 (s, 1H, H-3); ¹³C NMR (D₂O) δ : 45.0 (C-5), 58.2 (C-8), 58.3 (C-9), 65.5 (C-7), 75.0 (C-6), 111.6 (C-1), 127.0 (C-3), 134.6 (C-7a); FAB MS: 185 ([M+H]⁺); HRMS: calcd for [M+H]⁺ (C₈H₁₃N₂O₃) 185.0926, found 185.0926

2: $[\alpha]_{D}$ –32 (*c* 0.35, MeOH); ¹H NMR (D₂O) δ : 1.86 (ddd, 1H, *J* 7.5, 8.75, 13.0 Hz, H-7), 2.32 (ddd, 1H, *J* 3.0, 5.75, 13.0 Hz, H-7), 3.78 (dd, 1H, *J* 4.25, 12.25 Hz, H-9), 3.97 (dd, 1H, *J* 3.5, 12.25 Hz, H-9), 4.03 (ddd, 1H, *J* 3.5, 4.25, 6.5 Hz, H-5), 4.12 (ddd, 1H, *J* 3.0, 6.5, 8.75 Hz, H-6), 4.89 (1H, dd, *J* 5.75, 7.5 Hz, H-8), 6.95 (s, 1H, H-1), 7.73 (s, 1H, H-3); ¹³C NMR (D₂O) δ : 35.9 (C-7), 60.2 (C-9), 60.3 (C-8), 61.0 (C-5), 64.0 (C-6), 124.0 (C-1), 131.2 (C-3), 143.0 (C-8a); FAB MS: 185 ([M+H]⁺); HRMS: calcd for [M+H]⁺ (C₈H₁₃N₂O₃) 185.0926, found 185.0923

3: $[\alpha]_{D}$ –17 (*c* 0.4, MeOH); ¹H NMR (D₂O) δ : 2.03 (ddd, 1H, J 10.5, 10.75, 12.5 Hz, H-8), 2.14 (ddd, 1H, J 3.0, 4.0, 12.5 Hz, H-8'), 3.99 (d, 1H, J 15.0 Hz, H-5), 4.06 (ddd, 1H, J 1.75, 4.0, 10.75 Hz, H-7), 4.15 (dd, 1H, J 1.75, 6.5 Hz, H-6), 4.37 (dd, 1H, J 6.5, 15.0 Hz, H-5'), 4.89 (dd, 1H, J 3.0, 10.5 Hz, H-9), 6.95 (s, 1H, H-1), 7.73 (s, 1H, H-3); ¹³C NMR (D₂O) δ: 35.8 (C-8), 45.3 (C-5), 61.6 (C-9), 66.9 (C-7), 69.4 (C-6), 120.6 (C-1), 126.8 (C-3), 134.1 (C-9a); FAB MS: 185 ([M+H]+); HRMS: calcd for [M+H]+ (C₈H₁₃N₂O₃) 185.0926, found 185.0929 **13**: $[\alpha]_{D}$ -82 (c 1.75, CHCl₃); ¹H NMR (CDCl₃) δ : 3.48 (bd, 1H, J 2.5 Hz, H-7), 3.60 (dd, 1H, J 3.25, 11.25 Hz, H-9), 3.85 (d, 1H, J 11.25 Hz, H-9'), 4.40 (d, 1H, J 3.25 Hz, H-5), 4.51, 4.73 (2d, 2H, J 12 Hz, OBn), 4.54, 4.62 (2d, 2H, J 11.5 Hz, OBn), 4.88 (d, 1H, J 2.5 Hz, H-8), 4.89 (s, 1H, H-6), 6.86 (s, 1H, H-1), 7.19-7.42 (m, 10H, arom.), 7.51 (s, 1H, H-3); ¹³C NMR (CDCl₃) δ : 43.5 (C-7), 59.2 (C-5), 63.5 (C-9), 69.5 (OCH₂Ph), 70.7 (OCH₂Ph), 85.2 (C-6), 98.6 (C-8), 121.2 (C-1), 128.0-129.0 (m, arom.), 131.1 (C-3), 135.2 (C-7a), 138.8, 138.9 (s-arom.); FAB MS: 363 ([M+H]⁺)

14: $[\alpha]_D$ +172 (*c* 0.9, CHCl₃); ¹H NMR (CDCl₃) δ : 3.47 (dd, 1H, *J* 8.75, 11.25 Hz, H-9), 3.60 (dd, 1H, *J* 5.0, 11.25 Hz, H-9), 4.24 (dd, 1H, *J* 1.25, 6.25 Hz, H-6), 4.41 (ddd, 1H, *J* 1.25, 5.0, 8.75 Hz, H-5), 4.42, 4.47 (2d, 2H, *J* 12.25 Hz, OBn), 4.88 (d, 1H, *J* 6.25, H-7), 4.95 (s, 2H, OBn), 6.94 (s, 1H, H-1), 7.20–7.40 (m, 10H, arom.), 7.50 (s, 1H, H-3); ¹³C NMR (CDCl₃) δ : 60.5 (C-5), 62.8 (C-9) 69.2 (OCH₂Ph), 69.3 (OCH₂Ph), 71.2 (C-6), 89.8 (C-7), 124.8 (C-1), 127.5– 129.0 (m, arom.), 138.3 (C-3), 139.2 (C-8a), 140.0 (C-8); FAB MS: 363 ([M+H]⁺) **15**: $[\alpha]_D$ +57 (*c* 1.5, CHCl₃); ¹H NMR (CDCl₃+C₆D₆) δ : 3.51 (dd, 1H, *J* 2.6, 13.2 Hz, H-5), 3.74 (ddd, 1H, *J* 2.6, 3.1, 9.0 Hz, H-6), 3.83 (dd, 1H, *J* 9.0, 13.2 Hz, H-5'), 3.88 (dd, 1H, *J* 3.1, 6.0 Hz, H-7), 4.21, 4.36 (2d, 2H, *J* 11.5 Hz, OBn), 4.58, 4.61 (2d, 2H, *J* 12 Hz, OBn), 4.70 (d, 1H, *J* 6.0, H-8), 7.00 (s, 1H, H-1), 7.00–7.30 (m, 10H, arom.), 7.46 (s, 1H, H-3); ¹³C NMR (CDCl₃+C₆D₆) δ : 47.9 (C-5) 69.9 (C-6), 70.3 (OCH₂Ph), 71.2 (OCH₂Ph), 76.4 (C-7), 95.8 (C-8), 127.5–129.0 (m, arom.), 131.0 (C-3), 137.7, 139.4 (s-arom.), 138.0 (C-9a), 140.0 (C-1), 147.8 (C-9); FAB MS: 363 ([M+H]⁺).